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Received July 09, 2014; accepted for publication August 06, 2014; available online without subscription through the open access option.

©AlphaMed Press
1066-5099/2014/\$30.00/0

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/stem.1857

Concise Review: the Obesity Cancer Paradigm: Exploration of the Interactions and Cross-Talk with Adipose Stem Cells

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Key words. Adult stem cells • adipose stem cells • Breast cancer • Stromal cells • Cytokines • Cancer

ABSTRACT

With the recognition of obesity as a global health crisis, researchers have devoted greater effort to defining and understanding the pathophysiological molecular pathways regulating the biology of adipose tissue and obesity. Obesity, the excessive accumulation of adipose tissue due to hyperplasia and hypertrophy, has been linked to an increased incidence and aggressiveness of colon, hematological, prostate, and postmenopausal breast cancer cancers. The increased morbidity and mortality of obesity-associated cancers has been attributed to higher levels of hormones, adipokines, and cytokines secreted by the adipose tissue. The increased amount of adipose tissue also results in higher numbers of adipose stromal/stem cells (ASCs). These ASCs have been shown to impact cancer progression directly through several mechanisms, including the increased recruitment of ASCs to the tumor site and increased production of cytokines and growth factors by ASCs and other cells within the tumor stroma. Emerging evidence indicates that obesity induces alterations in the biologic properties of ASCs, subsequently leading to enhanced tumorigenesis and metastasis of cancer cells. This review will discuss the links between obesity and cancer tumor progression, including obesity-associated changes in adipose tissue, inflammation, adipokines and chemokines. Novel topics will include a discussion of the contribution of ASCs to this complex system with an emphasis on their role in the tumor stroma. The reciprocal and circular feedback loop between obesity and ASCs as well as the mechanisms by which ASCs from obese patients alter the biology of cancer cells and enhance tumorigenesis will be discussed. STEM CELLS 2014; 00:000–000

INTRODUCTION

More than one third of adults in the United States are obese, which is a number that has increased significantly in the last 10 years [1]. According to the World Health Organization statistics, obesity rates across the globe have almost doubled since 1980. The distinction between being overweight and obese is determined by the body mass index (BMI), calculated based on the height and weight of an individual. An individual with a BMI of 24.9 to 29.9 is considered overweight, while a person with a BMI greater than 30.0 is defined as ob-

ese. On a global scale, 1.4 billion adults meet the requirements for being overweight and nearly 500 million adults meet the requirements for being obese worldwide [2].

In 2007, the World Cancer Research Fund employed meta-analytic procedures to study the effects of obesity on cancer incidence and mortality. They found that higher levels of adiposity were associated with increased rates of colorectal, postmenopausal breast, and renal carcinomas [3]. Furthermore, additional meta-analysis confirmed an association between obesity and several other cancers in both men and women, includ-

ing endometrial, prostate, and esophageal cancers, malignant melanoma, hematological malignancies and large B-cell lymphomas [4-13]. Clearly, a better understanding of the mechanism(s) by which obesity enhances tumorigenesis is both a necessity and a priority.

Types of Adipose Tissue and their Role in Obesity

Historically, endocrinologists have divided adipose tissue into two categories, white adipose tissue (WAT) or brown adipose tissue (BAT). WAT is further subdivided into unique depots based on the location and its function: visceral (*around the organs*) and subcutaneous (*between the muscle and the dermal fascia*). The visceral WAT stores excess energy but also provides physical protection to the organs. For instance, perirenal fat is superficial to the renal capsule and protects the kidney from trauma. In contrast, the primary function of subcutaneous WAT is to store excess triglycerides and release free fatty acids during extended periods of fasting, starvation, or exercise. It has also been suggested that subcutaneous WAT functions as a buffer during intake of dietary lipids to protect the organs against the lipotoxicity of free fatty acid oxidation [14].

In contrast, BAT oxidizes chemical energy to produce heat, through the actions of mitochondrial uncoupling protein-1 (UCP1), as a defense against hypothermia [15]. Human babies, who lack body hair or a protective coat, have significant brown fat depots, presumably to provide heat in the cold environment encountered following birth. As humans age, BAT levels decrease. However, recent studies have identified an additional type of adipose tissue that is a hybrid between white adipose tissue and brown adipose tissue, termed beige or brite (***brown/white***) adipose tissue. Adults who have been exposed to chronic cold conditions form brown fat-like depots characterized by enhanced thermogenesis located in the supraclavicular and neck region [16-21]. These brown fat-like depots maintain high levels of expression of UCP1 and appear morphologically similar to brown fat. These brown fat-like depots have been located in regions where white adipose depots are generally found [22, 23]. Unlike classical BAT, which is derived from a myogenic factor 5 (Myf5) muscle-like cellular lineage, the beige/brite adipocytes lack Myf5 expression [24].

While all adipose depot sites can increase in volume, only an accumulation of WAT increases the risk of developing various diseases, including heart disease, cancer, metabolic syndrome, and stroke [25-28]. Extensive reviews have focused on the association of obesity with heart disease, metabolic syndrome, and stroke [29-35]. The focus of this review will be on the relationship between increased adiposity, the biology of adipose stromal/stem cells (ASCs), and tumorigenesis.

Adipose Tissue and Adipose Stromal/Stem Cells (ASCs)

Once considered solely as an energy reservoir or thermal insulator, adipose tissue is now being recognized as a complex endocrine organ involved in energy homeostasis, feeding, reproduction, and inflammation. Adipose tissue is heterogeneous, containing adipocytes and cells from the stromal vascular fraction, namely ASCs (15-30%), endothelial cells (10-20%), pericytes (3-5%), granulocytes (10-15%), monocytes (5-15%), and lymphocytes (10-15%) [36].

Among the cell types within the stromal vascular fraction, ASCs have recently been the focus of research because they have the potential to differentiate into mesenchymal tissue such as osteocytes, chondrocytes, and adipocytes, are immune privileged and have immunomodulatory properties. Because they do not express MHC class II molecules or costimulatory molecules [37, 38], ASCs are immune privileged. ASCs have a complex biology in regards to their anti-inflammatory properties; these cells inhibit natural killer cell activation, resulting in impaired cytotoxicity processes [37]. ASCs reduce the proliferation of B cells, reduce immunoglobulin production, and suppress B cell functions [39]. These features make ASCs ideal for tissue engineering and regenerative medicine, since these cells have the potential to differentiate into many cell types and immunomodulate the immune system without causing rejection by the host or the grafted cells [40-45].

Obesity Related Alterations to Adipose Tissue and the Impact on Cancer

Obesity alters the physiological function of adipose tissue, resulting in chronic inflammation, skewed secretion of adipokines, and changes to the biology of ASCs. Adipose tissue expansion in obesity increases the distance between the enlarging adipocytes and their vasculature, leading to localized hypoxia. Adipocytes can grow up to 100-200 μm in diameter and subsequently exceed the typical diffusion distances of oxygen into tissue [46, 47]. The oxygen content in expanded adipose tissue is close to zero at 100 μm distances from the vasculature, implying that increased adipocyte size and adipocyte number results in significant hypoxia [47]. Furthermore, other studies have shown that despite the substantial increase in adipose tissue associated with obesity, neither cardiac output nor total blood flow to the adipose tissue is increased [48, 49]. In obese mice, the reduced blood perfusion and hypoxia appear to be specific to WAT [50]. The lack of oxygen to the adipose tissue results in the activation of hypoxia-induced factor 1-alpha (HIF-1 α) and increased angiogenesis; however, the response is insufficient to compensate for the growing adipocytes, which leads to chronic low-grade inflammation [51, 52]. It is postulated that this chronic low-grade inflammation induces the excess secretion of pro-inflammatory cytokines, chemokines, protease, and protease inhibitors, such as tumor necrosis factor-alpha

(TNF- α), interleukin 6 (IL-6), monocyte chemoattractant protein 1 (MCP-1), leptin, and plasminogen activator inhibitor type 1 (PAI-1), which lead to adipose tissue dysfunction [53, 54]. The role that each of these factors plays in obesity and cancer will be presented in more detail.

TNF- α . TNF- α has an important role in the adaptive response of the immune system and other organ systems. TNF- α is an endogenous pyrogen that can induce fever, apoptotic cell death, inflammation as well as inhibiting tumorigenesis. However, dysregulation of TNF- α has been implicated in a variety of human diseases, including cancer, because it activates the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) pathway, leading to the expression of a variety of inflammation-related genes [55, 56]. TNF- α appears to contribute to the development of the tissue architecture necessary for tumor growth and metastasis [57]. It has also been shown to induce the production of other cytokines, angiogenic factors and matrix metalloproteinases (MMPs), which may drive the survival and metastasis of tumor cells [58]. Furthermore, long-term exposure of hormone receptor positive breast cancer cells to TNF- α induces an epithelial-to-mesenchymal transition (EMT), a process by which tumor cells lose their cell-to-cell adhesion and gain migratory properties that facilitate metastasis [59].

IL-6. Similarly, IL-6 is an important regulator of immune cell growth and differentiation. Recent studies demonstrate that IL-6 regulates chronic inflammation, which can create a cellular microenvironment conducive to cancer growth [60]. High concentrations of circulating IL-6 in obese patients correlate with an increased risk of developing tumors. The production of the IL-6 receptor/ligand complex activates both Janus kinase (JAK) and the signal transducer and activator of transcription 3 (STAT3) pathways, which are key regulators of cell proliferation and apoptosis.

MCP-1. MCP-1 has been shown to recruit macrophages in both obesity and cancer [61, 62]. MCP-1 levels in adipose tissue and plasma are increased in genetically obese diabetic (db/db) mice and in wild-type mice fed a high fat diet [63]. With respect to cancer, stromal MCP-1 is involved in both tumor progression and metastasis [64]. Treatment of immunodeficient mice bearing human breast cancer cells with a neutralizing antibody to MCP-1 resulted in a significant reduction in macrophage infiltration, angiogenic activity, and overall tumor volume [64].

Leptin. In an obese state, leptin resistance causes hyperphagia, increased adipose tissue volume and hyperleptinemia, as the body attempts to compensate for the resistance [65-67]; however, increasing leptin secretion is ineffective. In fact, it has been shown that the plasma concentration and mRNA expression of leptin in adipose tissue are directly related to the severity of obesity [68, 69]. Hyperleptinemia is also partially responsible for the chronic low-grade inflammation associated with obesity. Excess leptin results in enhanced T-cell and macrophage activation as immune

cells respond to the leptin in the microenvironment. Leptin also increases the expression of TNF- α , reactive oxygen-species (ROS) production, MCP-1 expression, and endothelial cell proliferation and migration. These factors all increase cancer cell growth and mobility.

PAI-1. PAI-1 is a serine protease inhibitor (serpin) produced by many different cell types, including endothelial cells, stromal cells, and adipocytes. PAI-1 affects adipocyte differentiation and the expression of PAI-1 increases with higher levels of adiposity [70]. PAI-1 principally inhibits urokinase plasminogen activator (uPA), which acts as an inducer of fibrinolysis and extracellular matrix degradation [71]. PAI-1 expression is also associated with increased tumor cell invasion and metastasis [72], and some studies have shown that PAI-1 is a poor prognostic indicator for a number of cancers, including breast cancer and colon cancer [72, 73].

While most of the studies to date have focused on adipose tissue as a whole, few studies have investigated the impact of obesity on the ASCs. Due to the chronic low-grade inflammation within microenvironment of the adipose tissue, the biology of the ASCs within these depots may be altered. Studies have shown that obesity diminishes ASC differentiation potential along adipogenic and osteogenic lineages, indicating a possible reduction in stem cell properties in cells conditioned by obese environments [74, 75]. Other studies have indicated that ASCs from obese individuals promote luminal breast cancer cell proliferation, angiogenesis, and metastasis [76-78].

ASCs in the Tumor Stroma

The tumor stroma is composed of numerous cell types (immune system cells, fibroblasts, myofibroblasts, and vascular cells). One of the key cell types is the cancer-associated fibroblast (CAF). The number of CAFs increases with the aggressiveness of the cancer [79-82]. CAFs demonstrate similar characteristics as myofibroblasts and express alpha-smooth muscle actin (α -SMA), tenascin-C, nestin, neural/glial antigen 2 (NG2), and PDGFR- α [83, 84]. It has been shown that ASCs are recruited to the tumor, transition into CAFs, and then integrate into the stroma [85-87]. Recent data indicates that ASCs that have been exposed to cancer cells or tumor cell conditioned media express tenascin-C and α -SMA, which are characteristic of CAFs, and may provide some insights into their role in the tumor stroma [87].

The recruited ASCs can also stimulate tumor growth, promote angiogenesis, and increase cancer cell invasion [88-90]. When ASCs are exposed to exosomes from breast cancer cells, they increase the expression of tumor-promoting factors, such as stromal cell-derived factor 1 (SDF-1), vascular endothelial growth factor (VEGF), chemokine ligand 5 (CCL5), platelet-derived growth factor D (PDGF-D), and transforming growth factor beta (TGF- β) [85-87, 91-93]. This phenomenon correlated with the increased expression of TGF- β receptors and phosphorylation of key factors in the TGF- β receptor-mediated SMAD pathway in ASCs [85, 86].

Consequently, these ASCs promote cancer cell growth and stimulate metastasis [94]. *In vivo* studies have confirmed that simultaneous co-injection of primary breast cancer and ASCs into nude mice results in integration of ASCs into the tumor stroma, thereby increasing tumor volume and increasing the vascularity of the tumor [95-97].

Other studies have demonstrated that ASCs stimulate invasion and metastasis of cancer cells. Recent evidence demonstrated that ASCs enhanced the migration of several types of cancer: breast, colon, prostate, gastric, and head and neck tumors [95, 98-101]. Data from Muehlberg and colleagues indicated that implanting spheroids formed with breast cancer cells and ASCs into nude mice increased the number of lung metastases [102]. Together, these studies suggest that cancer cells can recruit ASCs to the tumor microenvironment, which in turn increases cancer cell proliferation and metastasis.

Mechanisms of ASC induced alterations in cancer cells and tumorigenesis

Breast cancer. While many studies have described the interaction between ASCs and breast cancer cells, only recently have studies extensively explored the mechanism by which this interaction occurs. ASCs stimulated by cancer cells secrete a wide range of cytokine, chemokines, and growth factors that, in turn, increase the proliferation of breast cancer cells in an ASC/cancer cell reciprocal feedback loop (Figure 1) [74]. More specifically, cancer cells activate ASCs to secrete SDF-1, which then binds to its receptor CXCR4 on breast cancer cells and induces cellular proliferation through protein kinase B (AKT), extracellular signal-regulated kinases 1/2 (ERK1/2), and Janus kinase- signal transducer and activator of transcription 3 (JAK2-STAT3) [102]. Potter and colleagues showed that ASCs induced the expression of chemokine (C-C motif) ligand 2 (CCL2), ETS domain-containing protein (ELK1), Ezrin (VIL2), and MMP-11 in primary epithelial cells and breast cancer cell lines, leading to increased tumor volume, neoangiogenesis, and epithelial cell migration [103].

A primary role for ASCs in the microenvironment is their ability to induce EMT and promote metastasis. Devarajan et al. found that ASC conditioned media induced expression of fibronectin, α -SMA, and vimentin in breast cancer cells, which are markers of EMT [91]. These results correlated with increased expansion of CD44^{high}/CD24^{low} cancer stem cells and anchorage-independent growth of cancer cells, leading to EMT of cancer cells [91]. Furthermore, Pinilla and colleagues described the association between CCL5 secretion by ASCs and elevated levels of MMP-9 activity within the tumor microenvironment, leading to increased tumor invasion. ASC-derived IL-6 and IL-8 have also been shown to increase migration, invasion, and anchorage-independent growth of breast cancer cell lines, including MDA-MB-231, T47D, and MCF7 cells [84, 100].

Colorectal cancer. While limited information on the effects of ASCs on colorectal cancer cells exists, studies have provided some insights on the interactions between ASCs and colorectal cancer cell proliferation, neoangiogenesis, and efficacy of chemotherapy agents. ASCs that underwent conversion to CAFs have been shown to release a variety of growth factors and cytokines, including SDF-1, IL-6 and VEGF that enhance the growth of colorectal cancer cells (Figure 1)[104-106]. Similar to breast cancer cells, SDF-1 elicits its effects through activation of CXCR4. This SDF-1/CXCR4 axis regulates phosphoinositide 3-kinase (PI3K/AKT), mitogen-activated protein kinase (MAPK), and uPA cascades, which ultimately alters chemotaxis, angiogenesis, and tumor metastasis in colorectal cancer cells [104-106]. Additional cytokines and chemokines secreted by ASCs into the tumor microenvironment increase the survival of the cancer cells [107]. For example, studies have demonstrated that ASCs secrete sufficient VEGF and IL-6 to induce neoangiogenesis, which is necessary to provide sufficient nutrients to the growing tumor [108]. Inhibition of VEGF or IL-6 leads to reduced angiogenesis and inhibition of tumor growth [109].

ASCs can also induce chemoresistance in colorectal cancer cells. These cells have been shown to become activated during treatment with platinum analogs and secrete factors that protect tumor cells against a variety of chemotherapeutic drugs [110, 111]. Distinct platinum-induced polyunsaturated fatty acids in minute quantities induced cancer cell resistance to a broad spectrum of chemotherapeutic agents [111]. Additional studies suggest that the secretion of interleukin 17 (IL-17) from ASCs, in response to chemotherapeutic agents, leads to chemoresistance and thus increases the number of colorectal cancer cells [112].

Prostate cancer. In prostate cancer, ASCs have been implicated in altering the gene expression profile of cancer cells, inducing a more aggressive phenotype, and increasing angiogenesis within the tumor (Figure 1) [92]. The number of ASCs was increased in cancer patients compared to prostatic nodular hyperplasia patients [99]. The ASCs are converted into CAFs and provide nutrients and support for the growing tumor. Ribeiro and colleagues found that adipose tissue and ASCs exposed to conditioned media from PC3 cells (prostate cancer cell line) had an altered adipokine expression profile, including increased osteopontin, TNF- α , and IL-6 [113]. These factors have been implicated in prostate cancer tumorigenicity and metastasis [114-117]. Prostate cancer cells co-injected with ASCs into nude mice caused increased tumor volume. The local delivery of oncostatin M exacerbated the effect of ASCs on prostate cancer cell proliferation and tumor volumes doubled in size [118]. Other studies have shown that ASCs mediate their effects via the SDF-1/CXCR4 axis. ASC-secreted SDF-1 increases the levels of CXCR4 that result in a more aggressive prostate cancer cell phenotype [101, 119]. ASCs have also been shown to increase capillary density as evidenced by increased expression

of VEGF, basic fibroblast growth factor (FGF2) and CD31 [101, 120]. There is emerging evidence that suggests ASCs primed with prostate cancer conditioned media can undergo neoplastic transformation, and these ASCs form prostate-like neoplastic lesions *in vivo* and produce aggressive tumors upon serial transplantation [121]. Additional studies will be necessary to determine the precise mechanism by which these primed ASCs undergo neoplastic transformation.

Obesity induced alterations to ASCs

Studies have shown that ASCs isolated from obese women have an increased potential to traffic to the tumor compared to the ASCs isolated from lean women [77]. Furthermore, studies investigating the impact of obesity on ASC have observed increase recruitment of ASCs to the tumor in obese, resulting in an increase in the number of circulating ASCs [77, 122]. Zhang and colleagues revealed that a higher number of ASCs could be isolated from the WAT of obese mice compared to lean mice, possibly due to increased volume of WAT in obese mice [122]. These studies have shown that once localized to the tumor microenvironment, the mobilized ASCs enhanced the tumor vasculature by transdifferentiation into perivascular cells and incorporating into the tumor microenvironment [122]. With more ASCs recruited to the tumor site in obese mice, the perivascular cells are able to provide oxygen and nutrients to the tumor, enhancing survival and limiting apoptosis of cancer cells (Figure 2) [122]. Consistent with Zhang et al., Bellows and colleagues found increased frequency of ASCs in the circulation of obese patients, compared to lean patients [123, 124].

Additional studies have shown that ASCs from obese women (obese ASCs) enhanced the proliferation of breast cancer cells *in vitro* (Figure 2) [78]. Interestingly, this phenomenon was restricted to ER⁺ breast cancer cells, suggesting that ASCs may act through an estrogen-mediated pathway [78]. These obese ASCs also express higher levels of leptin when they are stimulated with estrogen, suggesting an estrogen-mediated leptin response [78]. Inhibiting leptin expression using a leptin neutralizing antibody reduced the impact of obese ASCs on breast cancer cell proliferation *in vitro* [78]. Furthermore, obese ASCs have been shown to alter the expression of several key regulatory genes involved in the cell cycle, apoptosis, angiogenesis, EMT, and metastasis [78]. The expressions of these molecular markers in breast cancer are associated with poorer prognosis due to increased invasion and metastasis of breast cancer cells to distant organs [125-129]. These studies suggest the source of leptin within the microenvironment is the ASCs, and robust secretion of leptin by ASCs can promote cancer cell growth and progression.

Delivery of leptin to cancer cells either *in vitro* or *in vivo* has also demonstrated increased proliferation, migration, invasion, angiogenesis, and metastasis of the cells [130-132]. Pre-neoplastic colon epithelial cells exposed to leptin upregulated VEGF expression, resulting in

VEGF-driven angiogenesis and vascular development [133]. In breast cancer cells, leptin functions through the JAK2-STAT3, PI3K-AKT, ERK1/2, and activator protein 1 (AP-1) pathways, increasing the expression of proteolytic enzymes that are required in tumor growth, metastasis and neoangiogenesis [134-136]. In estrogen receptor-positive human breast cancer cell lines, leptin has been shown to exert its influence through the activation of the MAPK pathway [136]. Thus, high levels of leptin resulting from obesity may result in increased breast cancer incidence. In addition, future research on this topic should provide clues to the therapeutic potential of anti-leptin strategies.

CONCLUSIONS

Obesity is a major public health concern because it increases the risk of several debilitating and deadly diseases, including cancer [137]. While intense discussions on the mechanism(s) by which obesity impacts cancer are ongoing, recent studies suggest that ASCs, altered by obesity, integrate into the tumor stroma and provide support for the growing tumor. Numerous genes are differentially expressed in ASCs isolated from obese patients compared to those from lean patients. The data suggests that ASCs isolated from obese patients have an increased potential to assist cancer cells. Furthermore, the number of circulating ASCs in obese patients was significantly higher than in lean patients, which in turn may increase the opportunity for ASCs to home to tumors. Once recruited to the growing tumor, ASCs isolated from obese women not only produce a novel chemokine and cytokine repertoire but also express higher levels of chemokines and cytokines that further drive cancer cell proliferation and migration, tumor migration and invasion, and metastasis to distant organs.

While the body of literature presented in this review provides insight into our current understanding of the ASCs in the tumor stroma and the effects of obesity within this intricate microenvironment, further investigations are required. Future studies focused around the effects of obesity on ASCs and understand how obesity primes the ASCs resulting in increased tumorigenesis and/or metastasis will provide valuable insight to reducing cancer morbidity and mortality. Studies have also investigated the use of ASCs as vehicles for gene therapy and have gained significant attention [138-140]. Therefore, it is essential to identify the mechanism(s) by which ASCs influence cancer cells, since novel therapeutic targets can be developed to target ASCs and inhibit the growth and metastasis of cancer cells.

AUTHOR CONTRIBUTIONS

A.L.S.: Manuscript writing; M.E.B.: Final approval of manuscript; J.M.G.: Final approval of manuscript; B.A.B.: Final approval of manuscript

REFERENCES

- 1 James PT. Obesity: the worldwide epidemic. *Clin Dermatol.* 2004;22:276-280.
- 2 World Health Organization. **Global Health Observatory (GHO): Obesity.** Geneva, Switzerland; 2014.
- 3 Food, Nutrition, Physical Activity, and the Prevention of Cancer: a Global Perspective. In: Research AIFC, ed. 2nd ed. Washington, USA: WCRF. World Cancer Research Fund; 2007.
- 4 Bianchini F, Kaaks R, Vainio H. Overweight, obesity, and cancer risk. *Lancet Oncol.* 2002;3:565-574.
- 5 Buschemeyer WC, 3rd, Freedland SJ. Obesity and prostate cancer: epidemiology and clinical implications. *Eur Urol.* 2007;52:331-343.
- 6 Caldwell SH, Crespo DM, Kang HS et al. Obesity and hepatocellular carcinoma. *Gastroenterology.* 2004;127:S97-103.
- 7 Crosbie EJ, Roberts C, Qian W et al. Body mass index does not influence post-treatment survival in early stage endometrial cancer: results from the MRC ASTEC trial. *Eur J Cancer.* 2012;48:853-864.
- 8 Diaz ES, Karlan BY, Li AJ. Obesity-associated adipokines correlate with survival in epithelial ovarian cancer. *Gynecol Oncol.* 2013;129:353-357.
- 9 Fader AN, Arriba LN, Frasure HE et al. Endometrial cancer and obesity: epidemiology, biomarkers, prevention and survivorship. *Gynecol Oncol.* 2009;114:121-127.
- 10 Giovannucci E, Michaud D. The role of obesity and related metabolic disturbances in cancers of the colon, prostate, and pancreas. *Gastroenterology.* 2007;132:2208-2225.
- 11 Ma J, Li H, Giovannucci E et al. Prediagnostic body-mass index, plasma C-peptide concentration, and prostate cancer-specific mortality in men with prostate cancer: a long-term survival analysis. *Lancet Oncol.* 2008;9:1039-1047.
- 12 Reeves KW, Carter GC, Rodabough RJ et al. Obesity in relation to endometrial cancer risk and disease characteristics in the Women's Health Initiative. *Gynecol Oncol.* 2011;121:376-382.
- 13 Renehan AG, Tyson M, Egger M et al. Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet.* 2008;371:569-578.
- 14 Frayn KN. Adipose tissue as a buffer for daily lipid flux. *Diabetologia.* 2002;45:1201-1210.
- 15 Fedorenko A, Lishko PV, Kirichok Y. Mechanism of fatty-acid-dependent UCP1 uncoupling in brown fat mitochondria. *Cell.* 2012;151:400-413.
- 16 Huttunen P, Hirvonen J, Kinnula V. The occurrence of brown adipose tissue in outdoor workers. *Eur J Appl Physiol Occup Physiol.* 1981;46:339-345.
- 17 Cypess AM, Lehman S, Williams G et al. Identification and importance of brown adipose tissue in adult humans. *N Engl J Med.* 2009;360:1509-1517.
- 18 van Marken Lichtenbelt WD, Vanhomerig JW, Smulders NM et al. Cold-activated brown adipose tissue in healthy men. *N Engl J Med.* 2009;360:1500-1508.
- 19 Ahmadi N, Hajsadeghi F, Conneely M et al. Accurate detection of metabolically active "brown" and "white" adipose tissues with computed tomography. *Acad Radiol.* 2013;20:1443-1447.
- 20 Cypess AM, Doyle AN, Sass CA et al. Quantification of human and rodent brown adipose tissue function using 99mTc-methoxyisobutylisonitrile SPECT/CT and 18F-FDG PET/CT. *J Nucl Med.* 2013;54:1896-1901.
- 21 Reddy NL, Jones TA, Wayte SC et al. Identification of brown adipose tissue using MR imaging in a human adult with histological and immunohistochemical confirmation. *J Clin Endocrinol Metab.* 2014;99:E117-121.
- 22 Wu J, Boström P, Sparks LM et al. Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. *Cell.* 2012;150:366-376.
- 23 Ye L, Wu J, Cohen P et al. Fat cells directly sense temperature to activate thermogenesis. *Proc Natl Acad Sci U S A.* 2013;110:12480-12485.
- 24 Seale P, Bjork B, Yang W et al. PRDM16 controls a brown fat/skeletal muscle switch. *Nature.* 2008;454:961-967.
- 25 Guh DP, Zhang W, Bansback N et al. The incidence of co-morbidities related to obesity and overweight: a systematic review and meta-analysis. *BMC Public Health.* 2009;9:88.
- 26 Fogelholm M. Physical activity, fitness and fatness: relations to mortality, morbidity and disease risk factors. A systematic review. *Obes Rev.* 2010;11:202-221.
- 27 Perez-Hernandez AI, Catalan V, Gomez-Ambrosi J et al. Mechanisms Linking Excess Adiposity and Carcinogenesis Promotion. *Front Endocrinol.* 2014;5:65.
- 28 Doyle SL, Donohoe CL, Lysaght J et al. Visceral obesity, metabolic syndrome, insulin resistance and cancer. *Proc Nutr Soc.* 2012;71:181-189.
- 29 Grundy SM. Obesity, metabolic syndrome, and cardiovascular disease. *J Clin Endocrinol Metab.* 2004;89:2595-2600.
- 30 Jahangir E, De Schutter A, Lavie CJ. The relationship between obesity and coronary artery disease. *Transl Res.* 2014.
- 31 Lavie CJ, Milani RV, Ventura HO. Obesity and cardiovascular disease: risk factor, paradox, and impact of weight loss. *J Am Coll Cardiol.* 2009;53:1925-1932.
- 32 Lumeng CN, Saltiel AR. Inflammatory links between obesity and metabolic disease. *J Clin Invest.* 2011;121:2111-2117.
- 33 Poirier P, Giles TD, Bray GA et al. Obesity and cardiovascular disease: pathophysiology, evaluation, and effect of weight loss. *Arterioscler Thromb Vasc Biol.* 2006;26:968-976.
- 34 Reaven GM. Insulin resistance: the link between obesity and cardiovascular disease. *Med Clin North Am.* 2011;95:875-892.
- 35 Rosen ED, Spiegelman BM. What we talk about when we talk about fat. *Cell.* 2014;156:20-44.
- 36 Bourin P, Bunnell BA, Casteilla L et al. Stromal cells from the adipose tissue-derived stromal vascular fraction and culture expanded adipose tissue-derived stromal/stem cells: a joint statement of the International Federation for Adipose Therapeutics and Science (IFATS) and the International Society for Cellular Therapy (ISCT). *Cytotherapy.* 2013;15:641-648.
- 37 DelaRosa O, Sanchez-Correa B, Morgado S et al. Human adipose-derived stem cells impair natural killer cell function and exhibit low susceptibility to natural killer-mediated lysis. *Stem Cells Dev.* 2012;21:1333-1343.
- 38 Yanez R, Lamana ML, Garcia-Castro J et al. Adipose tissue-derived mesenchymal stem cells have in vivo immunosuppressive properties applicable for the control of the graft-versus-host disease. *Stem Cells.* 2006;24:2582-2591.
- 39 Bochev I, Elmadjian G, Kyurkchiev D et al. Mesenchymal stem cells from human bone marrow or adipose tissue differently modulate mitogen-stimulated B-cell immunoglobulin production in vitro. *Cell Biol Int.* 2008;32:384-393.
- 40 Gonzalez-Rey E, Gonzalez MA, Varela N et al. Human adipose-derived mesenchymal stem cells reduce inflammatory and T cell responses and induce regulatory T cells in vitro in rheumatoid arthritis. *Ann Rheum Dis.* 2010;69:241-248.
- 41 Semon JA, Maness C, Zhang X et al. Comparison of human adult stem cells from adipose tissue and bone marrow in the treatment of experimental autoimmune encephalomyelitis. *Stem Cell Res Ther.* 2014;5:2.
- 42 Semon JA, Zhang X, Pandey AC et al. Administration of murine stromal vascular fraction ameliorates chronic experimental autoimmune encephalomyelitis. *Stem Cells Transl Med.* 2013;2:789-796.
- 43 Zhang S, Danchuk SD, Bonvillain RW et al. Interleukin 6 mediates the therapeutic effects of adipose-derived stromal/stem cells in lipopolysaccharide-induced acute lung injury. *Stem Cells.* 2014.
- 44 Zhang S, Danchuk SD, Imhof KM et al. Comparison of the therapeutic effects of human and mouse adipose-derived stem cells in a murine model of lipopolysaccharide-induced acute lung injury. *Stem Cell Res Ther.* 2013;4:13.
- 45 Gimble JM, Katz AJ, Bunnell BA. Adipose-derived stem cells for regenerative medicine. *Circ Res.* 2007;100:1249-1260.

- 46 Brahimi-Horn MC, Pouyssegur J. Oxygen, a source of life and stress. **FEBS Lett.** 2007;581:3582-3591.
- 47 Folkman J, Hahnelfeldt P, Hlatky L. Cancer: looking outside the genome. **Nat Rev Mol Cell Biol.** 2000;1:76-79.
- 48 Blaak EE, van Baak MA, Kemerink GJ et al. Beta-adrenergic stimulation and abdominal subcutaneous fat blood flow in lean, obese, and reduced-obese subjects. **Metabolism.** 1995;44:183-187.
- 49 Jansson PA, Larsson A, Lonnroth PN. Relationship between blood pressure, metabolic variables and blood flow in obese subjects with or without non-insulin-dependent diabetes mellitus. **Eur J Clin Invest.** 1998;28:813-818.
- 50 West DB, Prinz WA, Francendese AA et al. Adipocyte blood flow is decreased in obese Zucker rats. **Am J Physiol.** 1987;253:R228-233.
- 51 Carroll VA, Ashcroft M. Targeting the molecular basis for tumour hypoxia. **Expert Rev Mol Med.** 2005;7:1-16.
- 52 Trayhurn P. Hypoxia and adipose tissue function and dysfunction in obesity. **Physiol Rev.** 2013;93:1-21.
- 53 Ouchi N, Parker JL, Lugus JJ et al. Adipokines in inflammation and metabolic disease. **Nat Rev Immunol.** 2011;11:85-97.
- 54 Prieto-Hontoria PL, Perez-Matute P, Fernandez-Galilea M et al. Role of obesity-associated dysfunctional adipose tissue in cancer: a molecular nutrition approach. **Biochim Biophys Acta.** 2011;1807:664-678.
- 55 Kulbe H, Thompson R, Wilson JL et al. The inflammatory cytokine tumor necrosis factor- α generates an autocrine tumor-promoting network in epithelial ovarian cancer cells. **Cancer Res.** 2007;67:585-592.
- 56 Tomita Y, Yang X, Ishida Y et al. Spontaneous regression of lung metastasis in the absence of tumor necrosis factor receptor p55. **Int J Cancer.** 2004;112:927-933.
- 57 Balkwill F. Tumour necrosis factor and cancer. **Nat Rev Cancer.** 2009;9:361-371.
- 58 Wu Y, Zhou BP. TNF- α /NF- κ B/Snail pathway in cancer cell migration and invasion. **Br J Cancer.** 2010;102:639-644.
- 59 Antoon JW, Lai R, Struckhoff AP et al. Altered death receptor signaling promotes epithelial-to-mesenchymal transition and acquired chemoresistance. **Sci Rep.** 2012;2:539.
- 60 Hodge DR, Hurt EM, Farrar WL. The role of IL-6 and STAT3 in inflammation and cancer. **Eur J Cancer.** 2005;41:2502-2512.
- 61 Li M, Knight DA, L AS et al. A role for CCL2 in both tumor progression and immunosurveillance. **Oncoimmunology.** 2013;2:e25474.
- 62 Panee J. Monocyte Chemoattractant Protein 1 (MCP-1) in obesity and diabetes. **Cytokine.** 2012;60:1-12.
- 63 Kanda H, Tateya S, Tamori Y et al. MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. **J Clin Invest.** 2006;116:1494-1505.
- 64 Fujimoto H, Sangai T, Ishii G et al. Stromal MCP-1 in mammary tumors induces tumor-associated macrophage infiltration and contributes to tumor progression. **Int J Cancer.** 2009;125:1276-1284.
- 65 Coppari R, Bjorbaek C. Leptin revisited: its mechanism of action and potential for treating diabetes. **Nat Rev Drug Discov.** 2012;11:692-708.
- 66 Liuzzi A, Savia G, Tagliaferri M et al. Serum leptin concentration in moderate and severe obesity: relationship with clinical, anthropometric and metabolic factors. **Int J Obes Relat Metab Disord.** 1999;23:1066-1073.
- 67 Vazquez-Vela ME, Torres N, Tovar AR. White adipose tissue as endocrine organ and its role in obesity. **Arch Med Res.** 2008;39:715-728.
- 68 Considine RV, Sinha MK, Heiman ML et al. Serum immunoreactive-leptin concentrations in normal-weight and obese humans. **N Engl J Med.** 1996;334:292-295.
- 69 Yang R, Barouch LA. Leptin signaling and obesity: cardiovascular consequences. **Circ Res.** 2007;101:545-559.
- 70 Tilg H, Moschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. **Nat Rev Immunol.** 2006;6:772-783.
- 71 Dass K, Ahmad A, Azmi AS et al. Evolving role of uPA/uPAR system in human cancers. **Cancer Treat Rev.** 2008;34:122-136.
- 72 Hogan NM, Joyce MR, Murphy JM et al. Impact of mesenchymal stem cell secreted PAI-1 on colon cancer cell migration and proliferation. **Biochem Biophys Res Commun.** 2013;435:574-579.
- 73 Sternlicht MD, Dunning AM, Moore DH et al. Prognostic value of PAI1 in invasive breast cancer: evidence that tumor-specific factors are more important than genetic variation in regulating PAI1 expression. **Cancer Epidemiol Biomarkers Prev.** 2006;15:2107-2114.
- 74 Kucerova L, Altanerova V, Matuskova M et al. Adipose tissue-derived human mesenchymal stem cells mediated prodrug cancer gene therapy. **Cancer Res.** 2007;67:6304-6313.
- 75 Perez LM, Bernal A, San Martin N et al. Metabolic rescue of obese adipose-derived stem cells by Lin28/Let7 pathway. **Diabetes.** 2013;62:2368-2379.
- 76 Orecchioni S, Gregato G, Martin-Padura I et al. Complementary populations of human adipose CD34+ progenitor cells promote growth, angiogenesis, and metastasis of breast cancer. **Cancer Res.** 2013;73:5880-5891.
- 77 Strong AL, Semon JA, Strong TA et al. Obesity-associated dysregulation of calpastatin and MMP-15 in adipose-derived stromal cells results in their enhanced invasion. **Stem Cells.** 2012;30:2774-2783.
- 78 Strong AL, Strong TA, Rhodes LV et al. Obesity associated alterations in the biology of adipose stem cells mediate enhanced tumorigenesis by estrogen dependent pathways. **Breast Cancer Res.** 2013;15:R102.
- 79 Shimoda M, Mellody KT, Orimo A. Carcinoma-associated fibroblasts are a rate-limiting determinant for tumour progression. **Semin Cell Dev Biol.** 2010;21:19-25.
- 80 Hasegawa T, Yashiro M, Nishii T et al. Cancer-associated fibroblasts might sustain the stemness of scirrhous gastric cancer cells via transforming growth factor- β signaling. **Int J Cancer.** 2014;134:1785-1795.
- 81 Orimo A, Gupta PB, Sgroi DC et al. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. **Cell.** 2005;121:335-348.
- 82 Mueller L, Goumas FA, Affeldt M et al. Stromal fibroblasts in colorectal liver metastases originate from resident fibroblasts and generate an inflammatory microenvironment. **Am J Pathol.** 2007;171:1608-1618.
- 83 Sugimoto H, Mundel TM, Kieran MW et al. Identification of fibroblast heterogeneity in the tumor microenvironment. **Cancer Biol Ther.** 2006;5:1640-1646.
- 84 Welte G, Alt E, Devarajan E et al. Interleukin-8 derived from local tissue-resident stromal cells promotes tumor cell invasion. **Mol Carcinog.** 2012;51:861-868.
- 85 Cho JA, Park H, Lim EH et al. Exosomes from ovarian cancer cells induce adipose tissue-derived mesenchymal stem cells to acquire the physical and functional characteristics of tumor-supporting myofibroblasts. **Gynecol Oncol.** 2011;123:379-386.
- 86 Cho JA, Park H, Lim EH et al. Exosomes from breast cancer cells can convert adipose tissue-derived mesenchymal stem cells into myofibroblast-like cells. **Int J Oncol.** 2012;40:130-138.
- 87 Jotzu C, Alt E, Welte G et al. Adipose tissue-derived stem cells differentiate into carcinoma-associated fibroblast-like cells under the influence of tumor-derived factors. **Anal Cell Pathol (Amst).** 2010;33:61-79.
- 88 De Wever O, Demetter P, Mareel M et al. Stromal myofibroblasts are drivers of invasive cancer growth. **Int J Cancer.** 2008;123:2229-2238.
- 89 Angelucci C, Maulucci G, Lama G et al. Epithelial-stromal interactions in human breast cancer: effects on adhesion, plasma membrane fluidity and migration speed and directness. **PLoS One.** 2012;7:e50804.
- 90 Zhao M, Sachs PC, Wang X et al. Mesenchymal stem cells in mammary adipose tissue stimulate progression of breast cancer resembling the basal-type. **Cancer Biol Ther.** 2012;13:782-792.
- 91 Devarajan E, Song YH, Krishnappa S et al. Epithelial-mesenchymal transition in breast cancer lines is mediated through PDGF-D released by tissue-resident stem cells. **Int J Cancer.** 2012;131:1023-1031.
- 92 Tuxhorn JA, Ayala GE, Smith MJ et al. Reactive stroma in human prostate

- cancer: induction of myofibroblast phenotype and extracellular matrix remodeling. **Clin Cancer Res.** 2002;8:2912-2923.
- 93 De Boeck A, Hendrix A, Maynard D et al. Differential secretome analysis of cancer-associated fibroblasts and bone marrow-derived precursors to identify microenvironmental regulators of colon cancer progression. **Proteomics.** 2013;13:379-388.
- 94 Pinilla S, Alt E, Abdul Khalek FJ et al. Tissue resident stem cells produce CCL5 under the influence of cancer cells and thereby promote breast cancer cell invasion. **Cancer Lett.** 2009;284:80-85.
- 95 Scherzed A, Hackenberg S, Radeloff A et al. Human mesenchymal stem cells promote cancer motility and cytokine secretion in vitro. **Cells Tissues Organs.** 2013;198:327-337.
- 96 Zimmerlin L, Donnenberg AD, Rubin JP et al. Regenerative therapy and cancer: in vitro and in vivo studies of the interaction between adipose-derived stem cells and breast cancer cells from clinical isolates. **Tissue Eng Part A.** 2011;17:93-106.
- 97 Belmar-Lopez C, Mendoza G, Oberg D et al. Tissue-derived mesenchymal stromal cells used as vehicles for anti-tumor therapy exert different in vivo effects on migration capacity and tumor growth. **BMC Med.** 2013;11:139.
- 98 Samarajeewa NU, Yang F, Docanto MM et al. HIF-1 α stimulates aromatase expression driven by prostaglandin E2 in breast adipose stroma. **Breast Cancer Res.** 2013;15:R30.
- 99 Ribeiro R, Monteiro C, Silvestre R et al. Human periprostatic white adipose tissue is rich in stromal progenitor cells and a potential source of prostate tumor stroma. **Exp Biol Med.** 2012;237:1155-1162.
- 100 Walter M, Liang S, Ghosh S et al. Interleukin 6 secreted from adipose stromal cells promotes migration and invasion of breast cancer cells. **Oncogene.** 2009;28:2745-2755.
- 101 Lin G, Yang R, Banie L et al. Effects of transplantation of adipose tissue-derived stem cells on prostate tumor. **Prostate.** 2010;70:1066-1073.
- 102 Muehlberg FL, Song YH, Krohn A et al. Tissue-resident stem cells promote breast cancer growth and metastasis. **Carcinogenesis.** 2009;30:589-597.
- 103 Potter SM, Dwyer RM, Hartmann MC et al. Influence of stromal-epithelial interactions on breast cancer in vitro and in vivo. **Breast Cancer Res Treat.** 2012;131:401-411.
- 104 Huang WS, Chin CC, Chen CN et al. Stromal cell-derived factor-1/CXC receptor 4 and beta1 integrin interaction regulates urokinase-type plasminogen activator expression in human colorectal cancer cells. **J Cell Physiol.** 2012;227:1114-1122.
- 105 Kollmar O, Rupertus K, Scheuer C et al. Stromal cell-derived factor-1 promotes cell migration and tumor growth of colorectal metastasis. **Neoplasia.** 2007;9:862-870.
- 106 Saigusa S, Toiyama Y, Tanaka K et al. Stromal CXCR4 and CXCL12 expression is associated with distant recurrence and poor prognosis in rectal cancer after chemoradiotherapy. **Ann Surg Oncol.** 2010;17:2051-2058.
- 107 Kilroy GE, Foster SJ, Wu X et al. Cytokine profile of human adipose-derived stem cells: expression of angiogenic, hematopoietic, and pro-inflammatory factors. **J Cell Physiol.** 2007;212:702-709.
- 108 Rehman J, Traktuev D, Li J et al. Secretion of angiogenic and antiapoptotic factors by human adipose stromal cells. **Circulation.** 2004;109:1292-1298.
- 109 Nagasaki T, Hara M, Nakanishi H et al. Interleukin-6 released by colon cancer-associated fibroblasts is critical for tumour angiogenesis: anti-interleukin-6 receptor antibody suppressed angiogenesis and inhibited tumour-stroma interaction. **Br J Cancer.** 2014;110:469-478.
- 110 Saigusa S, Toiyama Y, Tanaka K et al. Cancer-associated fibroblasts correlate with poor prognosis in rectal cancer after chemoradiotherapy. **Int J Oncol.** 2011;38:655-663.
- 111 Castells M, Thibault B, Delord JP et al. Implication of tumor microenvironment in chemoresistance: tumor-associated stromal cells protect tumor cells from cell death. **Int J Mol Sci.** 2012;13:9545-9571.
- 112 Lotti F, Jarrar AM, Pai RK et al. Chemotherapy activates cancer-associated fibroblasts to maintain colorectal cancer-initiating cells by IL-17A. **J Exp Med.** 2013;210:2851-2872.
- 113 Ribeiro R, Monteiro C, Cunha V et al. Human periprostatic adipose tissue promotes prostate cancer aggressiveness in vitro. **J Exp Clin Cancer Res.** 2012;31:32.
- 114 Zheng J, Hou ZB, Jiao NL. Effects of osteopontin downregulation on the growth of prostate cancer PC-3 cells. **Mol Med Rep.** 2011;4:1225-1231.
- 115 Liu H, Chen A, Guo F et al. A short-hairpin RNA targeting osteopontin downregulates MMP-2 and MMP-9 expressions in prostate cancer PC-3 cells. **Cancer Lett.** 2010;295:27-37.
- 116 Tse BW, Scott KF, Russell PJ. Paradoxical roles of tumour necrosis factor-alpha in prostate cancer biology. **Prostate Cancer.** 2012;2012:128965.
- 117 Michalaki V, Syrigos K, Charles P et al. Serum levels of IL-6 and TNF-alpha correlate with clinicopathological features and patient survival in patients with prostate cancer. **Br J Cancer.** 2004;90:2312-2316.
- 118 Lee MJ, Heo SC, Shin SH et al. Oncostatin M promotes mesenchymal stem cell-stimulated tumor growth through a paracrine mechanism involving periostin and TGFBI. **Int J Biochem Cell Biol.** 2013;45:1869-1877.
- 119 Darash-Yahana M, Pikarsky E, Abramovitch R et al. Role of high expression levels of CXCR4 in tumor growth, vascularization, and metastasis. **Faseb j.** 2004;18:1240-1242.
- 120 Prantl L, Muehlberg F, Navone NM et al. Adipose tissue-derived stem cells promote prostate tumor growth. **Prostate.** 2010;70:1709-1715.
- 121 Abd Elmaged ZY, Yang Y, Thomas R et al. Neoplastic reprogramming of patient-derived adipose stem cells by prostate cancer cell-associated exosomes. **Stem Cells.** 2014;32:983-997.
- 122 Zhang Y, Daquinag AC, Amaya-Manzanares F et al. Stromal progenitor cells from endogenous adipose tissue contribute to pericytes and adipocytes that populate the tumor microenvironment. **Cancer Res.** 2012;72:5198-5208.
- 123 Bellows CF, Zhang Y, Chen J et al. Circulation of Progenitor Cells in Obese and Lean Colorectal Cancer Patients. **Cancer Epidemiology Biomarkers & Prevention.** 2011;20:2461-2468.
- 124 Bellows CF, Zhang Y, Simmons PJ et al. Influence of BMI on level of circulating progenitor cells. **Obesity (Silver Spring).** 2011;19:1722-1726.
- 125 Hildenbrand R, Schaaf A. The urokinase-system in tumor tissue stroma of the breast and breast cancer cell invasion. **Int J Oncol.** 2009;34:15-23.
- 126 Mendes O, Kim HT, Lungu G et al. MMP2 role in breast cancer brain metastasis development and its regulation by TIMP2 and ERK1/2. **Clin Exp Metastasis.** 2007;24:341-351.
- 127 Nakopoulou L, Tsirmpa I, Alexandrou P et al. MMP-2 protein in invasive breast cancer and the impact of MMP-2/TIMP-2 phenotype on overall survival. **Breast Cancer Res Treat.** 2003;77:145-155.
- 128 Salgado R, Junius S, Benoy I et al. Circulating interleukin-6 predicts survival in patients with metastatic breast cancer. **Int J Cancer.** 2003;103:642-646.
- 129 Sullivan NJ, Sasser AK, Axel AE et al. Interleukin-6 induces an epithelial-mesenchymal transition phenotype in human breast cancer cells. **Oncogene.** 2009;28:2940-2947.
- 130 Cirillo D, Rachiglio AM, la Montagna R et al. Leptin signaling in breast cancer: an overview. **J Cell Biochem.** 2008;105:956-964.
- 131 Kim HS. Leptin and leptin receptor expression in breast cancer. **Cancer Res Treat.** 2009;41:155-163.
- 132 Amemori S, Ootani A, Aoki S et al. Adipocytes and preadipocytes promote the proliferation of colon cancer cells in vitro. **Am J Physiol Gastrointest Liver Physiol.** 2007;292:G923-929.
- 133 Birmingham JM, Busik JV, Hansen-Smith FM et al. Novel mechanism for obesity-induced colon cancer progression. **Carcinogenesis.** 2009;30:690-697.
- 134 Frankenberry KA, Skinner H, Somasundar P et al. Leptin receptor expression and cell signaling in breast cancer. **Int J Oncol.** 2006;28:985-993.
- 135 Gao J, Tian J, Lv Y et al. Leptin induces functional activation of cyclooxygenase-2 through JAK2/STAT3, MAPK/ERK, and PI3K/AKT pathways in human endometrial cancer cells. **Cancer Sci.** 2009;100:389-395.

- 136 Catalano S, Marsico S, Giordano C et al. Leptin Enhances, via AP-1, Expression of Aromatase in the MCF-7 Cell Line. **Journal of Biological Chemistry.** 2003;278:28668-28676.
- 137 De Pergola G, Silvestris F. Obesity as a major risk factor for cancer. **J Obes.** 2013;2013:291546.
- 138 Cavarretta IT, Altanerova V, Matuskova M et al. Adipose tissue-derived mesenchymal stem cells expressing prodrug-converting enzyme inhibit human prostate tumor growth. **Mol Ther.** 2010;18:223-231.
- 139 Grisendi G, Bussolari R, Cafarelli L et al. Adipose-derived mesenchymal stem cells as stable source of tumor necrosis factor-related apoptosis-inducing ligand delivery for cancer therapy. **Cancer Res.** 2010;70:3718-3729.
- 140 Kucerova L, Kovacovicova M, Polak S et al. Interaction of human adipose tissue-derived mesenchymal stromal cells with breast cancer cells. **Neoplasma.** 2011;58:361-370.

Figure 1. Model of the tumor-promoting effects of CAFs formed from ASCs. (A) Cancer cells secrete a wide range of cytokines, chemokines, and growth factors that play a role in the recruitment of several different cell types into the tumor. The tumor microenvironment is composed of cancer cells, endothelial cells, adipose stromal cells (ASCs), cancer associated fibroblasts (CAFs), and immune cells. (B) A reciprocal and circular feedback loop between cancer cells and ASCs is initiated by the secretion of cytokines from cancer cells. These cytokines activate ASCs, resulting in the conversion of ASCs into CAFs as noted by the increased expression in alpha-smooth muscle actin (α -SMA), tenascin-C, nestin, neuro-glial antigen 2, and platelet derived growth factor receptor-alpha (PDGFR- α). In turn, the CAFs secrete cytokines and chemokines that alter cancer cells, leading to an increase in the number of cancer cells, increased invasive potential of cancer cells, and potentially increased chemoresistance of cancer cells. (C) Cancer cells recruit ASCs into the microenvironment and induce their transformation into CAFs. This cellular conversion results in secretion of cytokines, chemokines, growth factors, and enzymes that enhance cancer cell proliferation, induce EMT and the metastasis of cancer cells to distant sites.

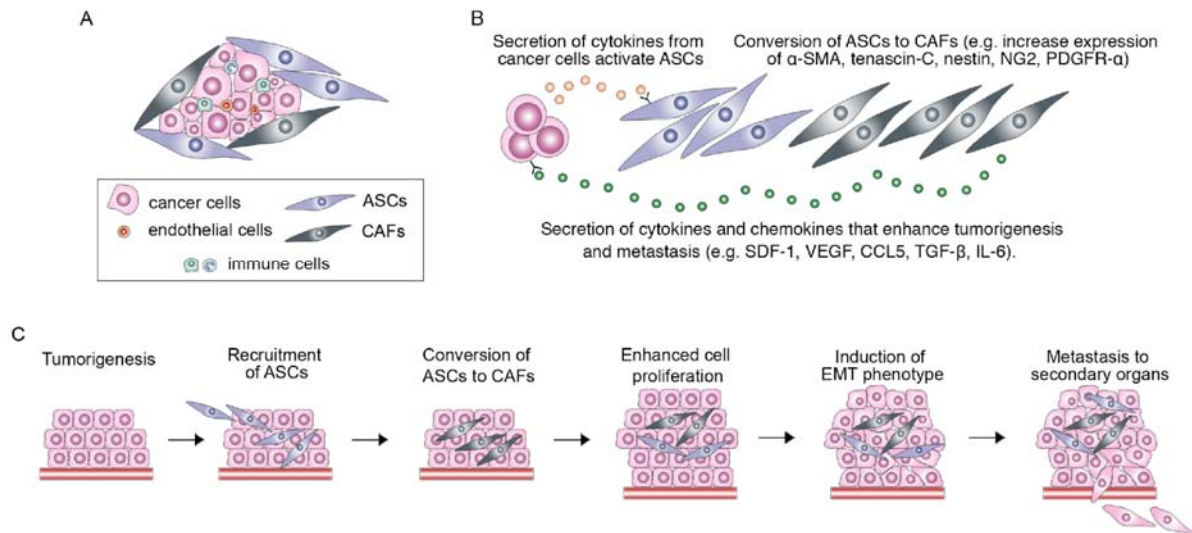


Figure 2. Model for the role of obesity in promoting tumorigenesis and cancer progression. The accumulation of adipose tissue in obese individuals, results in formation of an hypoxic environment surrounding adipocytes more distal to blood vessels. Consequently, the adipose tissue releases angiogenic factors that circulate through the vasculature to combat the hypoxia. The hypoxic environment also results in significant inflammation, which results in the secretion of pro-inflammatory cytokines. The secretion of pro-inflammatory cytokines within the adipose tissue microenvironment may, in turn, alter the tissue-resident stem cells (ASCs). The production of angiogenic factors, the secretion of inflammatory cytokines, and the perturbations to ASCs promote a microenvironment favorable for tumorigenesis and cancer progression.

